

CHEMICAL CONSTITUENTS FROM THE INFLORESCENCES OF *Ouratea hexasperma*

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In the course of our phytochemical investigation of Brazilian plants, we have studied some species of *Ouratea*, *Luxemburgia*, and *Salvagesia* (Ochnaceae). The studies of *Ouratea* allowed us to identify many classes of compounds besides the flavonoid dimers [1–4]. In the northeast of Brazil *Ouratea* species are used in folk medicine for several purposes with the trivial name “batiputa oil” [5]. The present work reports the first phytochemical study of *O. hexasperma* inflorescences and identifies three steroids, a saponin, and five known active glycopyranosyl flavonoids. The yellow inflorescences of *O. hexasperma* were collected in the Tabuleiro Region, Joao Pessoa-PB, Brazil, and a voucher sample was deposited at Herbarium Lauro Pires Xavier, Joao Pessoa-PB by the botanist Maria de Fatima Agra. The flowers (350.0 g) were dried, powdered, and extracted with methanol at room temperature. The solvent was removed under vacuum to yield a residue (50.0 g). The crude methanol residue was suspended in MeOH–H₂O (9:1) and successively partitioned with hexane and ethyl acetate. Each fraction was chromatographed on a silica gel column eluted with binary mixtures of hexane–ethyl acetate–methanol in increasing polarity gradients besides the pure solvents and Sephadex LH-20 to yield compounds 1–9. The compounds were investigated by spectroscopic methods, including NMR and mass spectrometry. The compounds were identified as campesterol (1), sitosterol (2), stigmasterol (3) [6], sitosterol β -D-glucopyranoside (4) [7, 8], rutin (5) [9], orientin (6) [10, 11], vitexin (7) [10, 11], swertisin (8) [12], and swertijaponin (9) [12].

Orientin (6). Yellow powder. IR (KBr, ν_{\max} , cm⁻¹): 3384, 3246 ($\nu_{\text{O-H}}$), 1655 ($\nu_{\text{C=O}}$), 1614, 1508, 1428 ($\delta_{\text{C-H}}$), 1094–1041 ($\nu_{\text{C-O}}$). ¹H NMR (200 MHz, DMSO-d₆, ppm, δ , J/Hz): 13.17 (s, 5-OH), 7.50 (1H, dd, $J_1 = 8.0$, $J_2 = 2.1$, H-6'), 7.44 (1H d, $J = 2.1$, H-2'), 6.90 (1H, d, $J = 8.2$, H-5'), 6.65 (1H, s, H-3), 6.25 (1H, s, H-6), 4.72 (1H, d, $J = 9.4$, H-1''), 3.82 (1H, t, $J = 9.4$, H-2''), 3.24 (1H, m, H-3''), 3.37 (1H, t, $J = 9.4$, H-4''), 3.21 (1H, m, H-5''), 3.75 (2H, m, H-6''). ¹³C NMR (50.3 MHz, DMSO-d₆, δ , ppm): 164.2 (C-2), 102.4 (C-3), 182.0 (C-4), 160.5 (C-5), 98.3 (C-6), 162.8 (C-7), 104.6 (C-8), 156.0 (C-9), 103.9 (C-10), 121.9 (C-1'), 114.1 (C-2'), 145.9 (C-3'), 149.9 (C-4'), 115.8 (C-5'), 119.4 (C-6'), 73.5 (C-1''), 70.9 (C-2''), 78.9 (C-3''), 70.8 (C-4''), 82.0 (C-5''), 61.8 (C-6'') [10, 11].

Vitexin (7). Yellow powder. ¹H NMR (200 MHz, DMSO-d₆, δ , ppm, J/Hz): 13.2 (s, 5-HO), 8.26 (2H, $J = 8.7$, H-2', 6'), 7.05 (2H, d, $J = 8.7$, H-3', 5'), 6.94 (1H, s, H-3), 6.44 (1H, s, H-6), 4.94 (1H, d, $J = 9.8$, H-1''), 3.82 (1H, t, $J = 9.6$, H-2''), 3.37 (1H t, $J = 9.4$, H-4''), 3.24 (1H m, H-3''), 3.21 (1H, m, H-5''), 3.75 (1H, br.d, $J = 11.6$, 2H-6''). ¹³C NMR (50.3 MHz, DMSO-d₆, δ , ppm): 164.9 (C-2), 102.5 (C-3), 182.7 (C-4), 160.3 (C-5), 98.4 (C-6), 162.3 (C-7), 104.6 (C-8), 155.6 (C-9), 104.1 (C-10), 122.1 (C-1'), 128.9 (C-2'), 115.0 (C-3'), 161.3 (C-4'), 115.0 (C-5'), 128.9 (C-6'), 73.9 (C-1''), 71.0 (C-2''), 79.0 (C-3''), 70.2 (C-4''), 81.2 (C-5''), 61.4 (C-6'') [10, 11].

Swertisin (8). Yellow amorphous solid. ¹H NMR (200 MHz, DMSO-d₆, δ , ppm, J/Hz): 13.17 (s, OH, H-5), 8.02 (2H, d, $J = 8.0$, H-2', 6'), 6.89 (2H, d, $J = 8.0$, H-3', 5'), 6.81 (1H, s, H-3), 6.51 (1H, s, H-8), 4.72 (1H, d, $J = 10.0$, H-1''), 3.82 (1H, m, H-2''), 3.25–3.75 (4H, m, H-3'', 4'', 5'', 6'') [12].

Swertijaponin (9). Yellow amorphous solid. ¹H NMR (200 MHz, DMSO-d₆, δ , ppm, J/Hz): 13.15 (s, OH, H-5), 7.56 (1H, br.d, $J = 8.5$, H-6'), 7.55 (1H, br.s, H-2'), 6.92 (1H, d, $J = 8.5$, H-5'), 6.79 (1H, s, H-3), 6.59 (1H, s, H-8), 4.76 (1H, d, $J = 10.0$, H-1''), 3.2–3.8 (5H, m, H-2''–6''), 3.86 (3H, s, OCH₃). ¹³C NMR (50.3 MHz, DMSO-d₆, δ , ppm): 164.5 (C-2),

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102.3 (C-3), 182.3 (C-4), 161.3 (C-5), 108.6 (C-6), 163.3 (C-7), 94.9 (C-8), 155.6 (C-9), 104.4 (C-10), 121.7 (C-1'), 113.9 (C-2'), 145.9 (C-3'), 150.1 (C-4'), 115.0 (C-5'), 119.5 (C-6'), 73.1 (C-1''), 70.7 (C-2''), 78.7 (C-3''), 70.7 (C-4''), 82.9 (C-5''), 61.6 (C-6''), 56.6 (OCH₃) [12].

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REFERENCES

1. J. R. Velandia, M. G. de Carvalho, R. Braz-Filho, and A. A. Werle, *Phytochem. Anal.*, **13**, 283 (2002).
2. I. C. Moreira, M. G. de Carvalho, A. B. F. O. Bastos, and R. Braz-Filho, *Phytochemistry*, **51**, 833 (1999).
3. I. C. Moreira, D. C. Sobrinho, M. G. de Carvalho, and R. Braz-Filho, *Phytochemistry*, **35**, 1567 (1994).
4. J. F. de S. Daniel, M. G. de Carvalho, R. da S. Cardoso, M. de F. Agra, and M. N. Eberlin, *J. Braz. Chem. Soc.*, **16**, 634 (2005).
5. G. M. Barroso, *Sistemática de Angiospermas do Brasil*, UFV-MG, Minas Gerais, 1986, 130 p.
6. M. R. G. Vega, M. G. de Carvalho, I. J. C. Vieira, and R. Braz-Filho, *J. Nat. Med.*, **62**, 122 (2008).
7. R. Braz Filho, H. E. Gottlieb, A. P. Mourao, and C. A. S. Miranda, *An. Acad. Bras. Ciências*, **58**, 363 (1986).
8. H. Kojima, N. Sato, A. Hatano, and H. Ogura, *Phytochemistry*, **29**, 2351 (1990).
9. P. K. Agrawal and M. C. Bansal, *Flavonoid Glycosides*, in: *Carbon-¹³NMR of Flavonoids*, Agrawal P. K. (ed.), Elsevier, Amsterdam, 1989, 564 p.
10. S. Rayyan, T. Fossen, S. H. Natelaand, and O. M. Anderse, *Phytochem. Anal.*, **19**, 334 (2005).
11. X. X. Zhou, J. Peng, G. Fan, and Y. Wu, *J. Chromatogr. A*, **1092**, 216 (2005).
12. Y. Kumarasamy, M. Byres, P. J. Cox, A. Delazar, M. Jaspars, M. Nahar, and S. D. Sarker, *Chem. Nat. Comp.*, **40**, 122 (2004).